

Brachyspirainfektioner hos svin og in situ identifikation

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Diagnostic Pathology

In situ detection methods

Detection of bacteria in formalin-fixed paraffin embedded tissue sections

- Classic, indicative stainings (e.g. Giemsa, Warthin-Starry)
- Specific stainings
 - immunohistochemistry, phenotype depending
 - *in situ* hybridization targeting ribosomal RNA, genotype depending

Fluorescent *in situ* hybridisation for identification of intestinal pathogens

Specific in situ probes

- *Domain bacterium*
- *Gammaproteobacteria*

- genus *Brachyspira*
- genus *Campylobacter*
- genus *Helicobacter*

Brachyspira

- *B. hyodysenteriae*
- *B. pilosicoli*
- *B. intermedia*
- *B. innocens*
- *B. murdochii*

- *Fusobacterium necrophorum*
- *Campylobacter jejuni/coli*
- *Clostridium perfringens*
- *Clostridium difficile*
- *Salmonella enterica*
- *Lawsonia intracellularis*

Intestinal diseases and differential diagnostic problems

- Proliferative enteropathy: *Lawsonia intracellularis*
- Post weaning diarrhoea / edema disease: *Escherichia coli*
- Swine dysentery : *Brachyspira hyodysenteriae*
- Spirochaetal colitis: *Brachyspira pilosicoli*
 - *Brachyspira intermedia*: pathogenic?
 - *Brachyspira innocens*: non-pathogenic?
 - *Brachyspira murdochii*: non-pathogenic?
 - "*Brachyspira suanatina*": colitis

- Salmonellosis
- Porcine Circovirus type 2 (PCV2)
- Other agents?

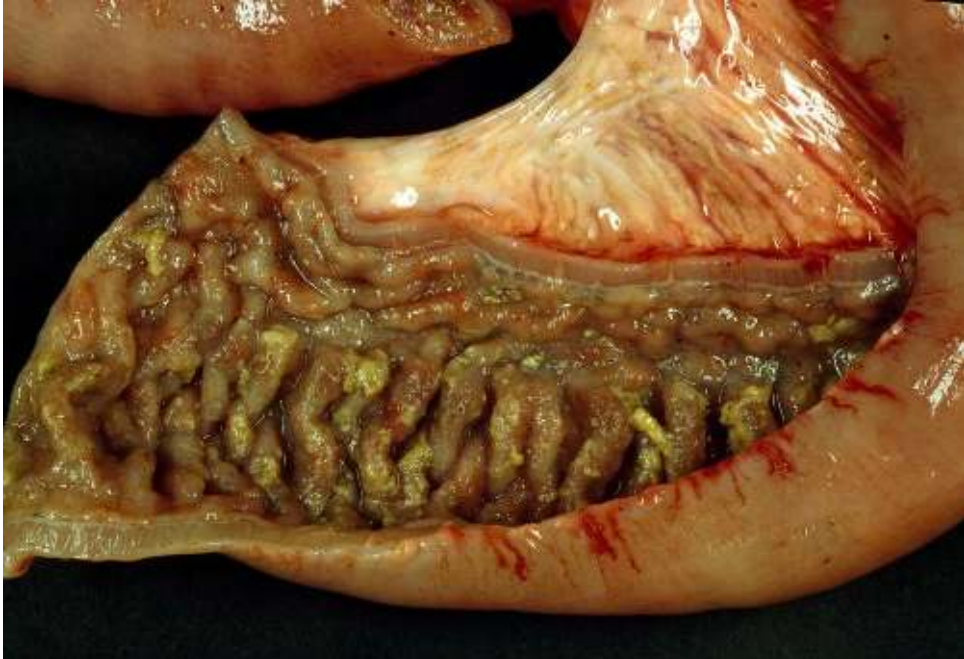
Svinedysenteri

- Smitsom mukohæmmorrhagisk colitis
- *Brachyspira hyodysenteriae*

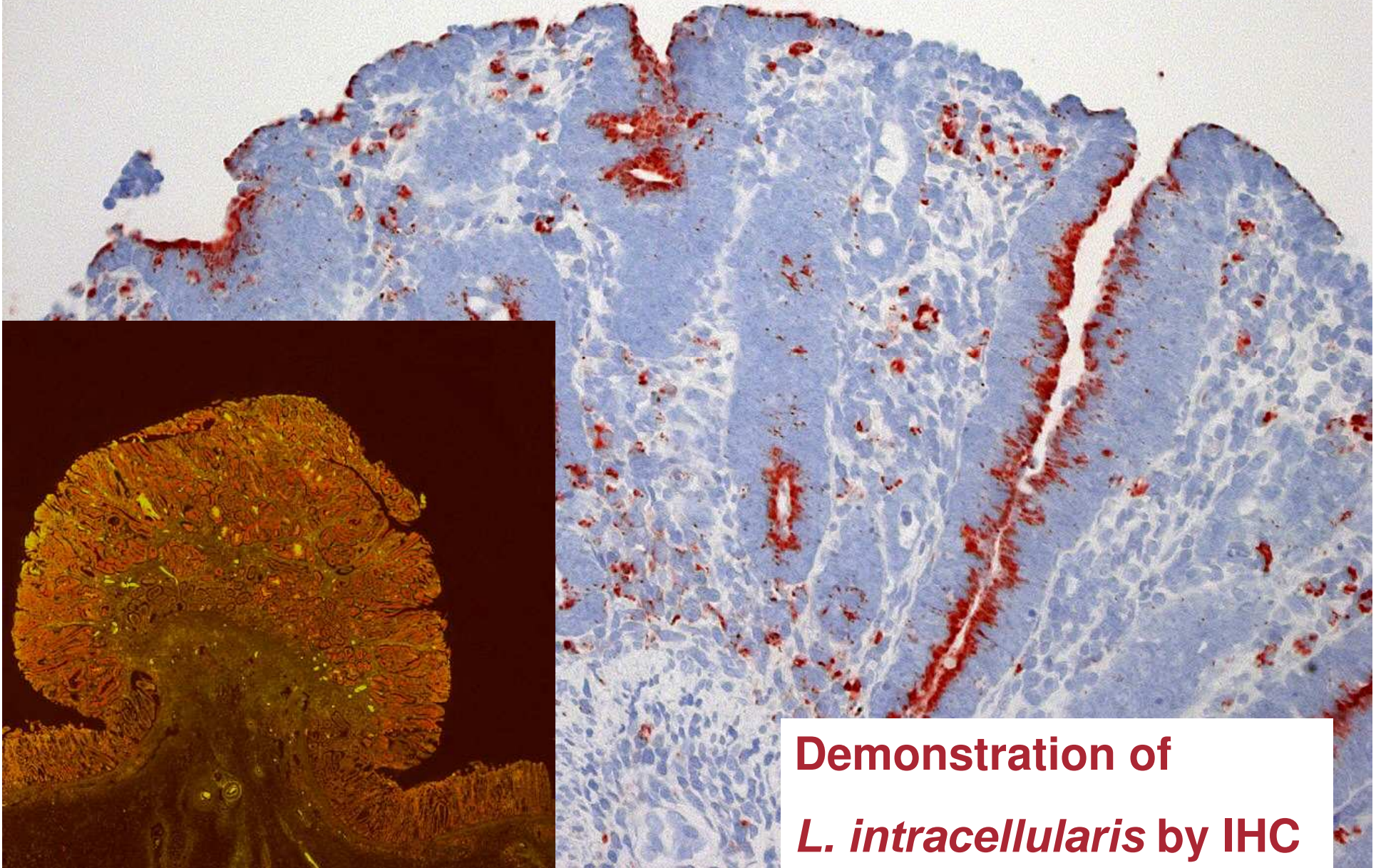
Andre Brachyspira

- *B. innocens*, non-patogen
- *B. intermedia*, patogen?
- *B. pilosicoli*, colitis, flere dyr. intestinal spirokæetose
- *B. murdochii*, non-patogen?
- ”*B. suanantina*”, kraftig hæmolyse, andefugle, svin (colitis)
- ”*B. hampsonii*”, kraftig hæmolyse, svin, colitis, nordamerika

Intestinal diseases and differential diagnostic problems

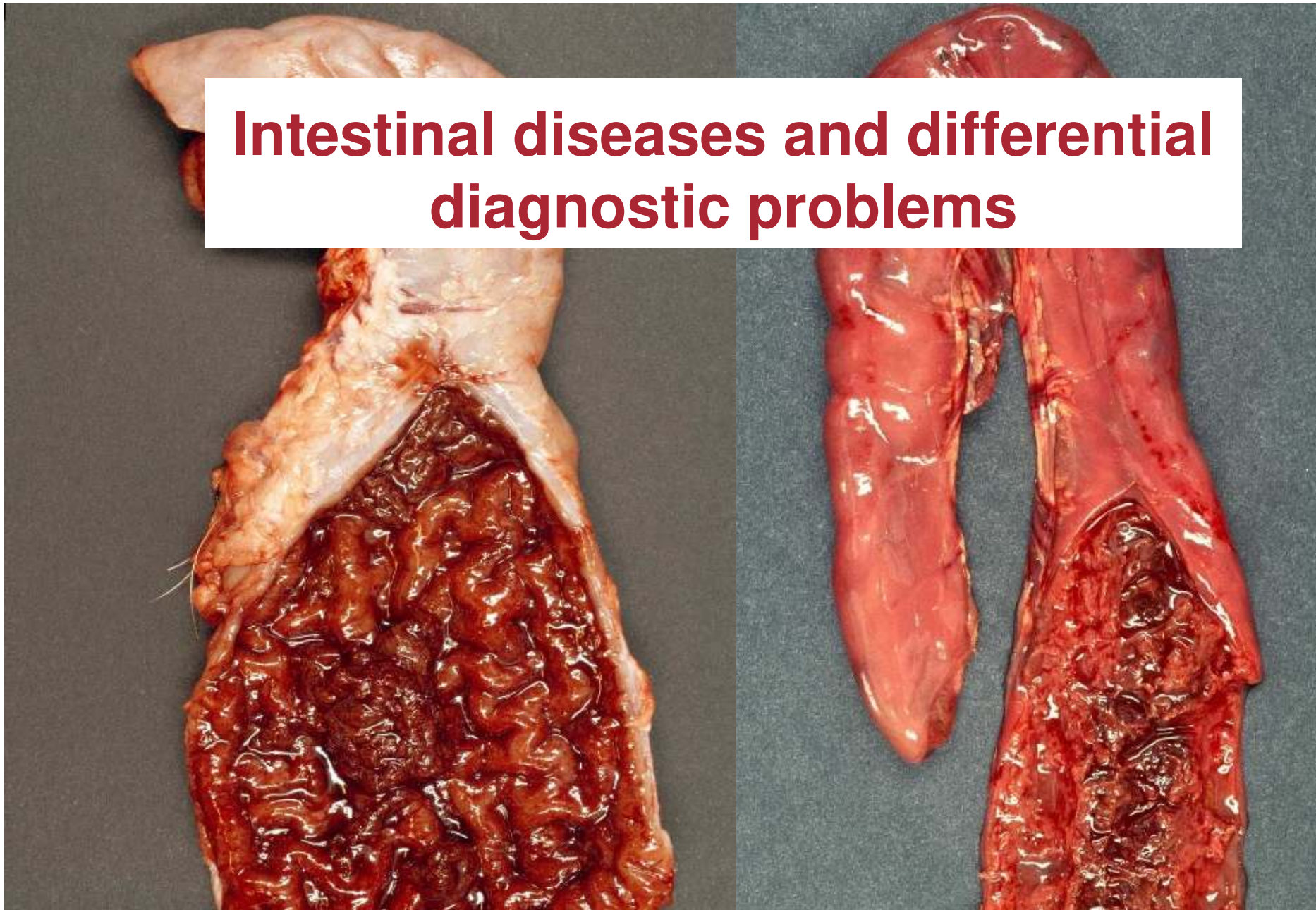


Proliferative enteropathy – *L. intracellularis*

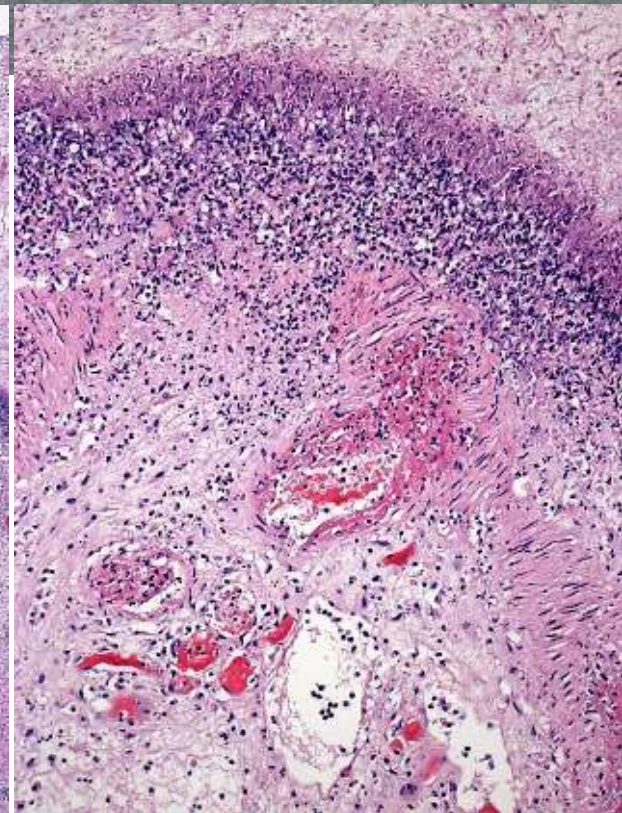
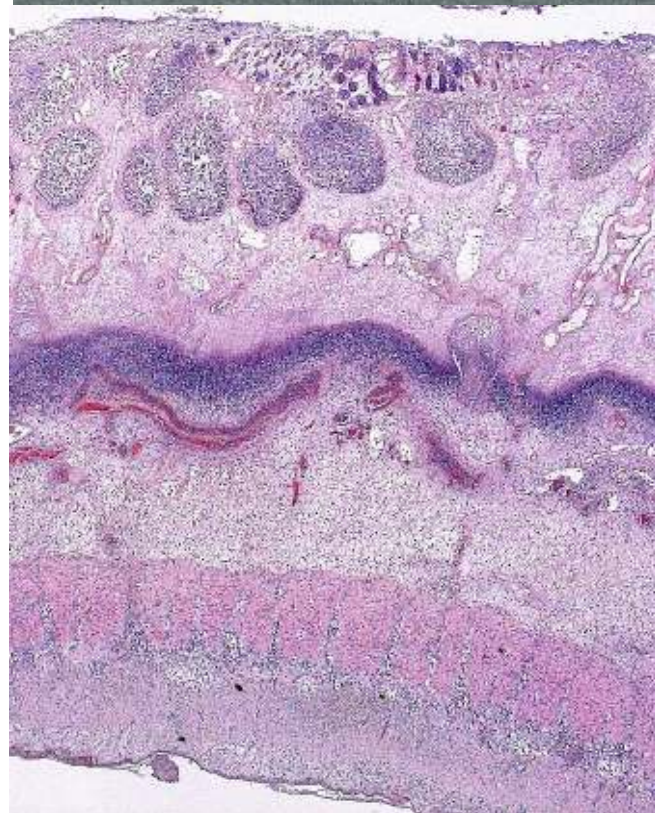


Demonstration of
L. intracellularis by IHC

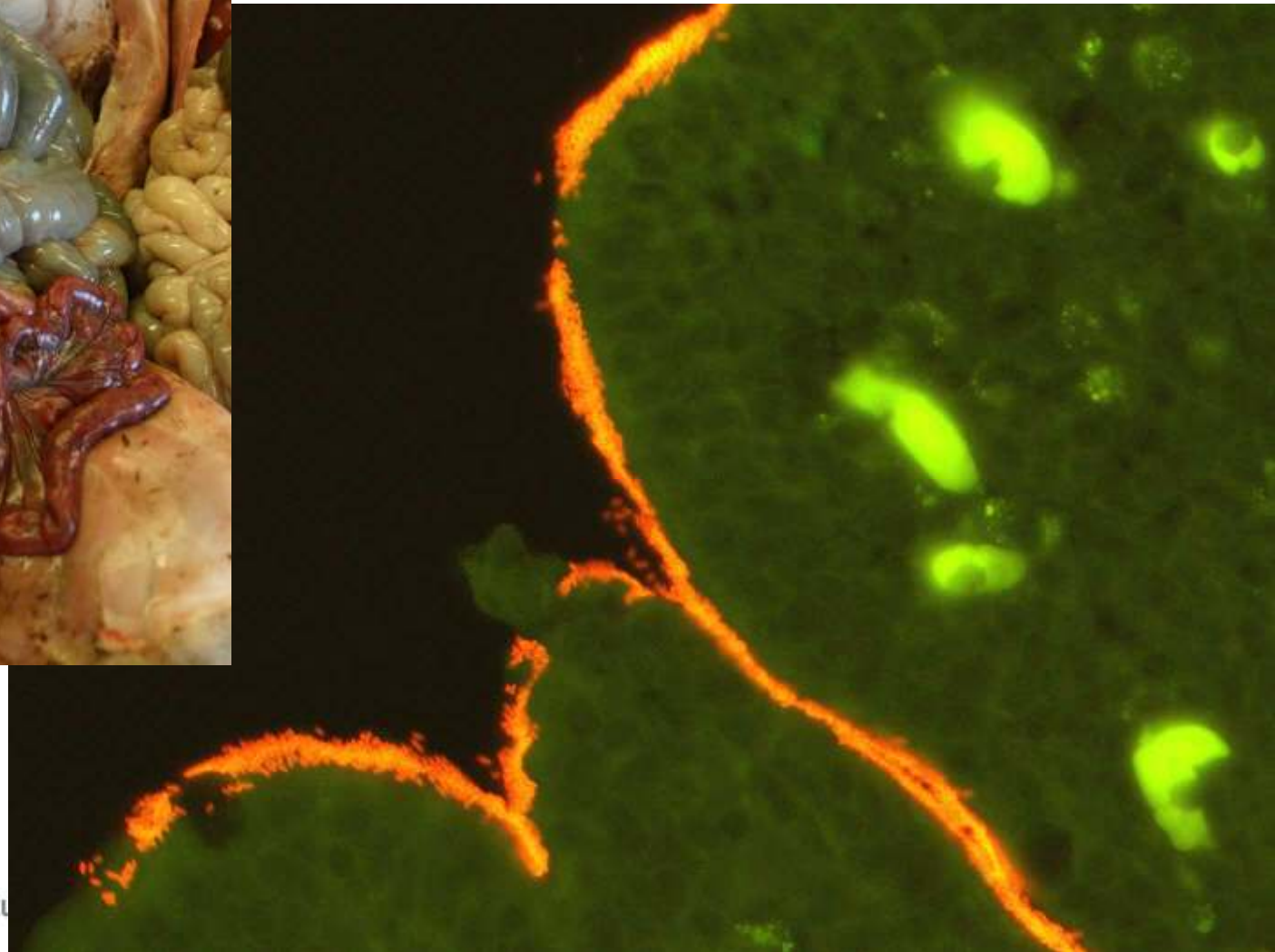
Intestinal diseases and differential diagnostic problems



Edema disease
***E. coli* positive for**
verotoxin 2e and
fimbria (F18).



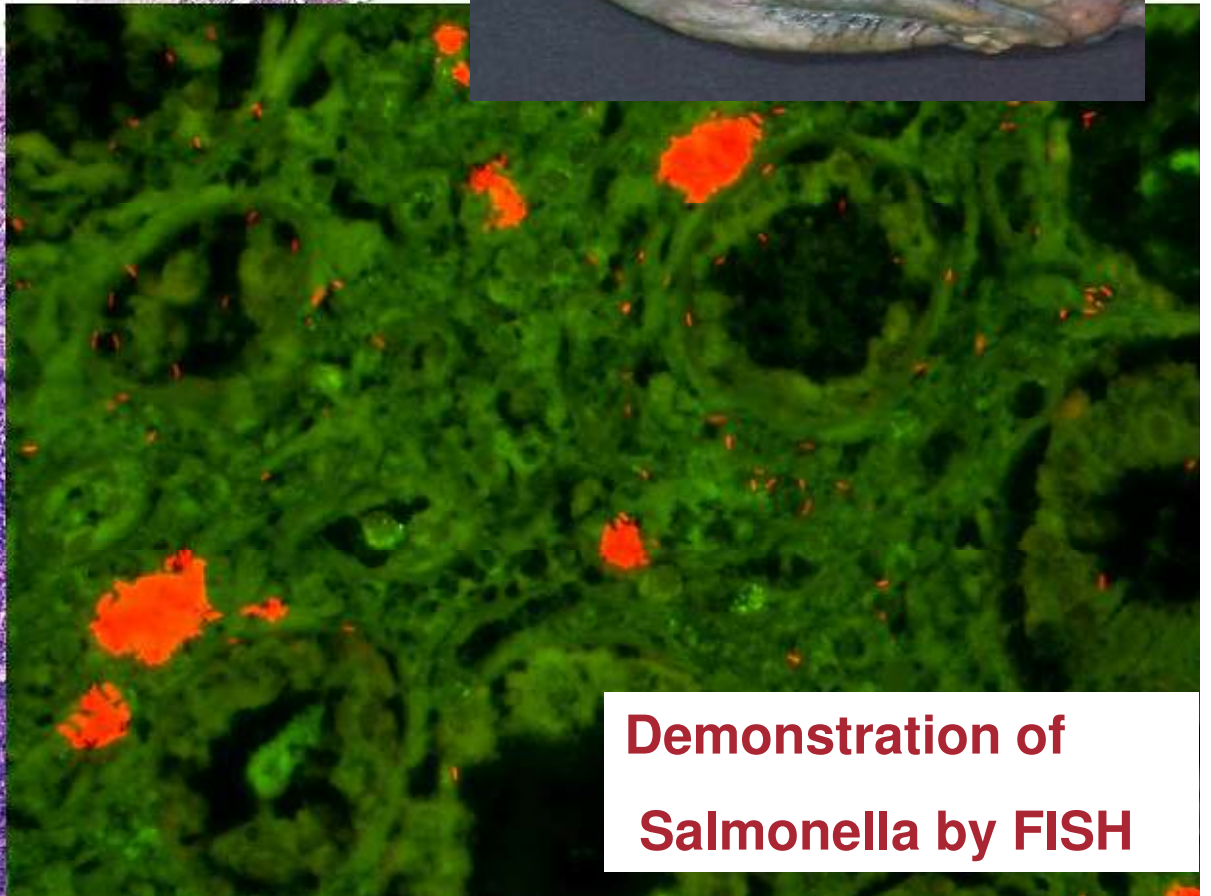
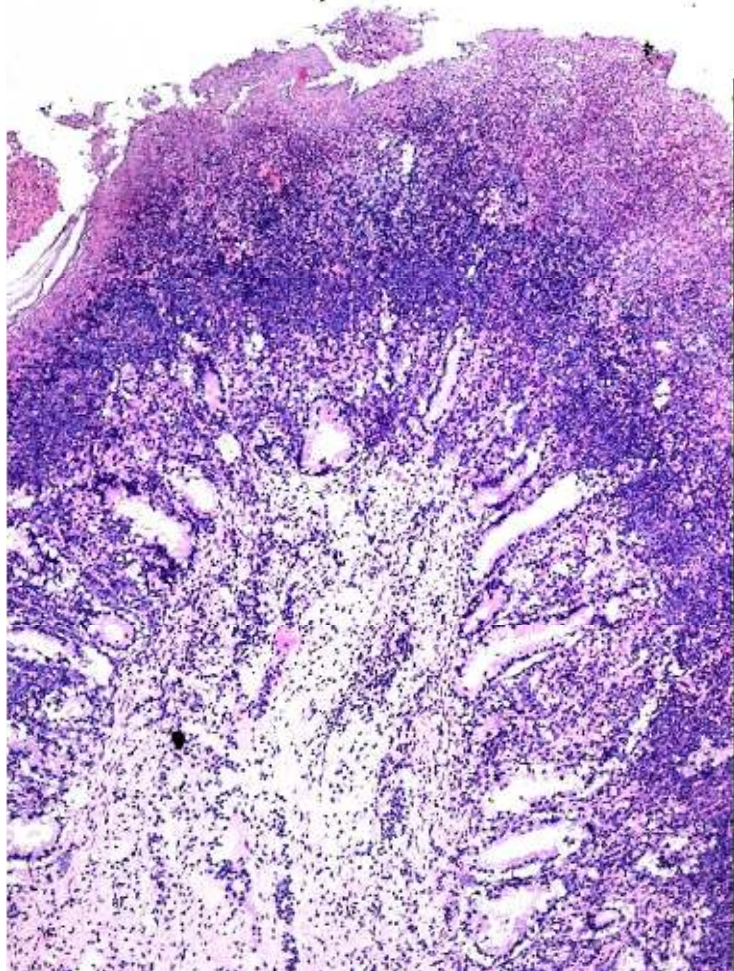
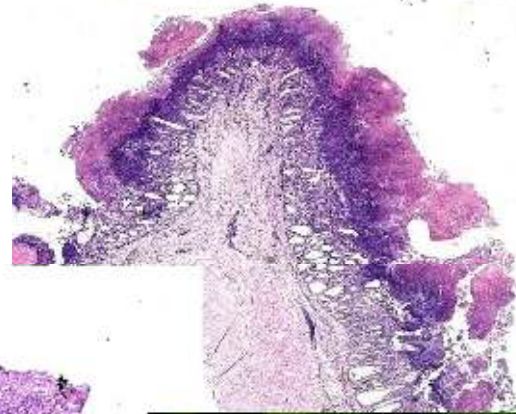
Demonstration of *E. coli* attached to villus epithelium in jejunum, FISH



Intestinal diseases and differential diagnostic problems

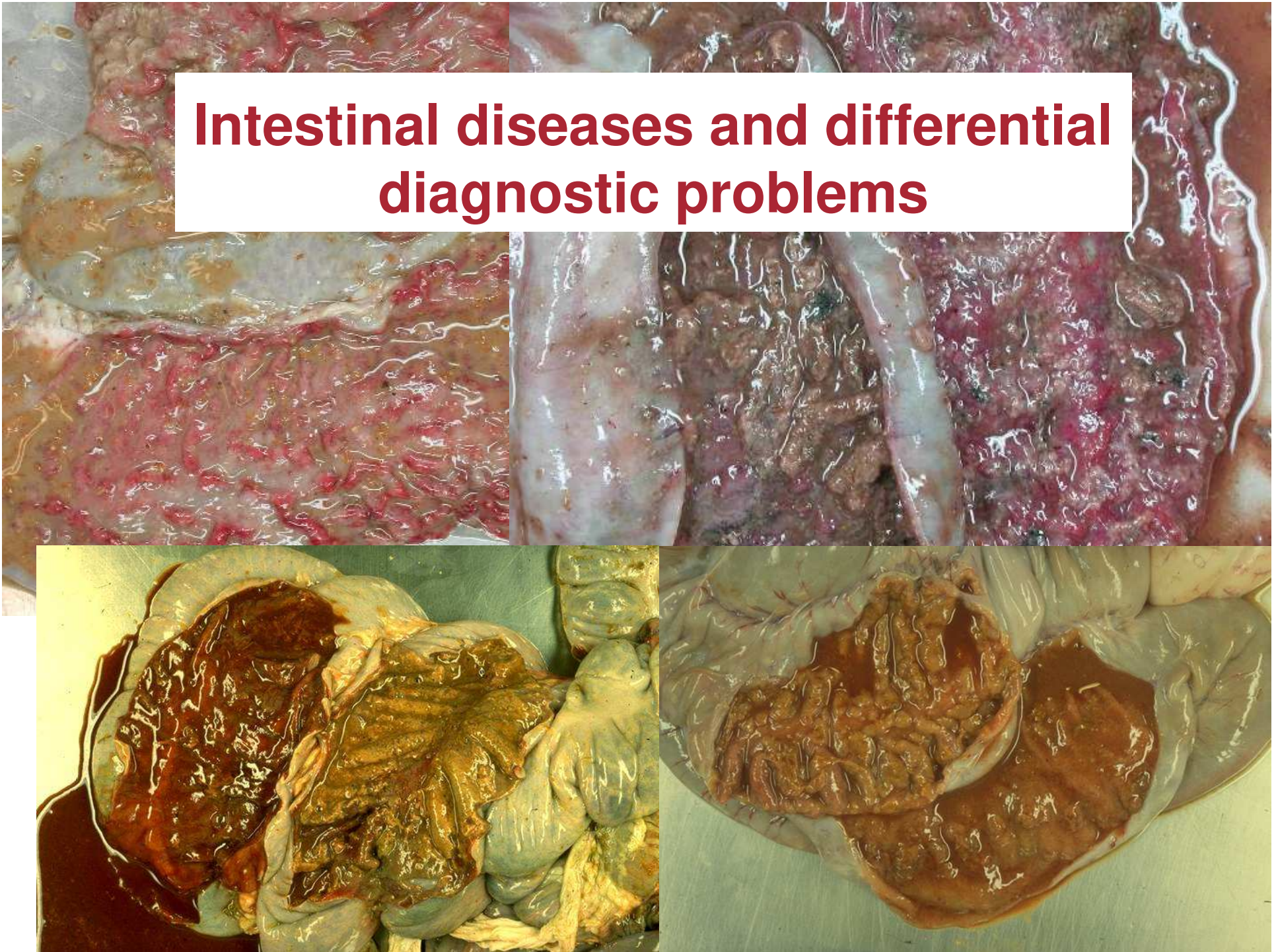


Salmonellosis

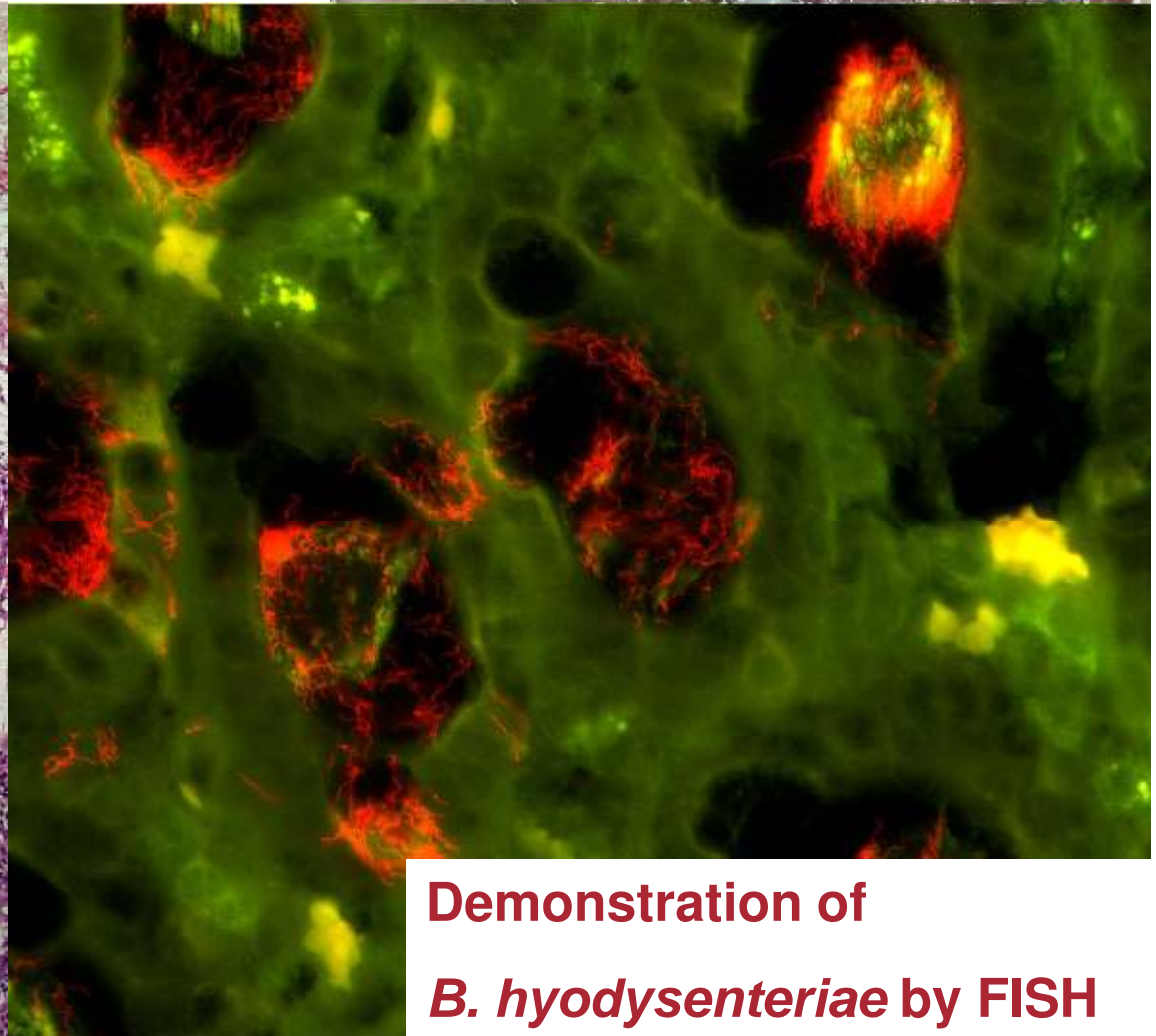
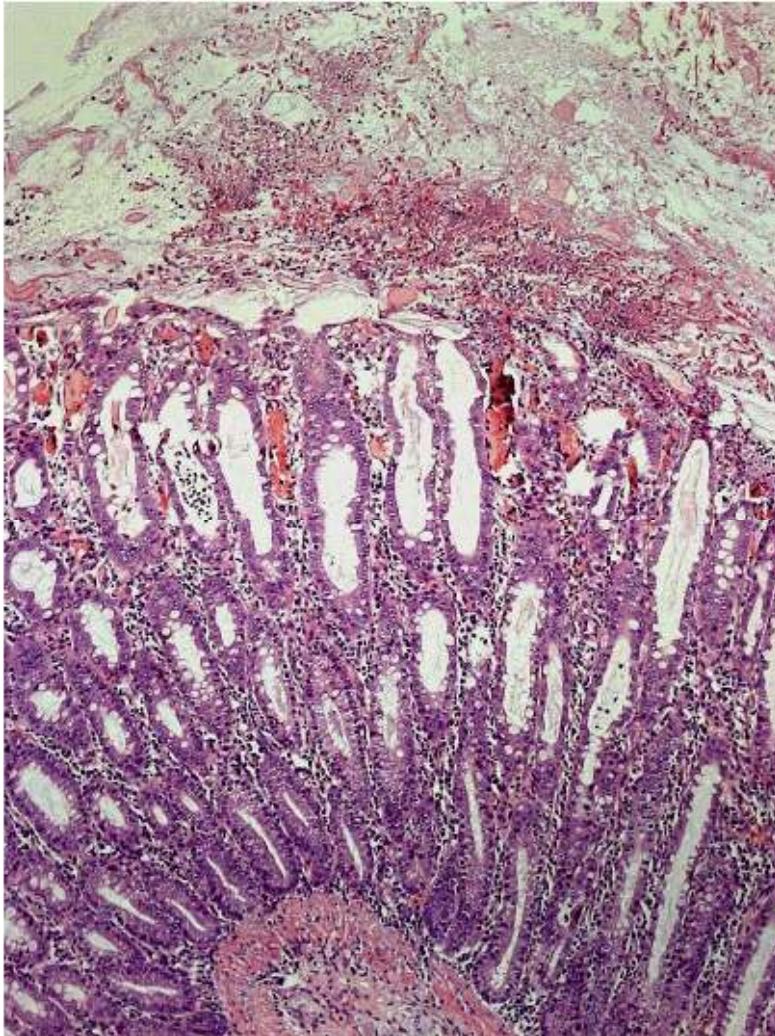


**Demonstration of
Salmonella by FISH**

Intestinal diseases and differential diagnostic problems



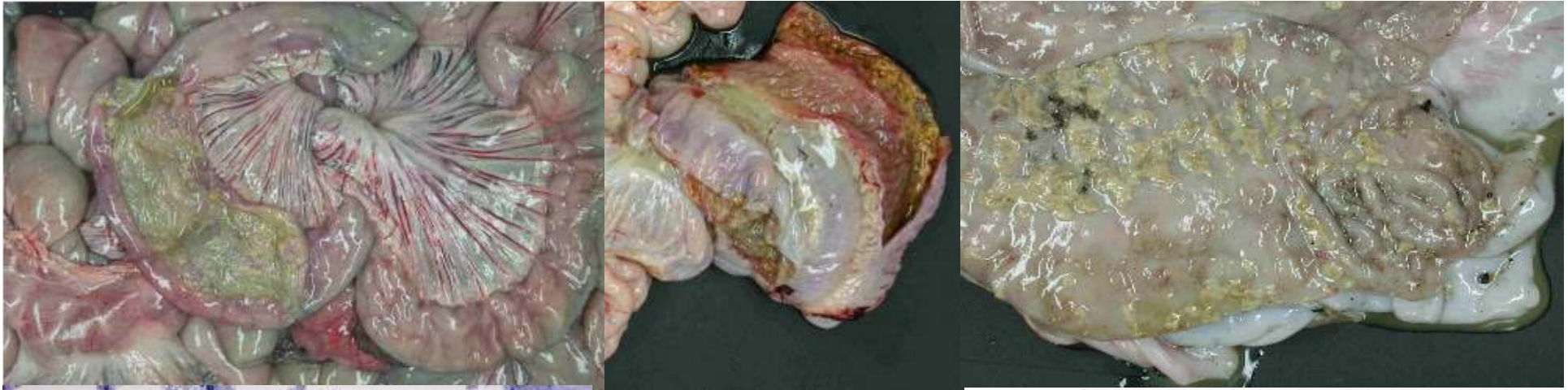
Swine dysentery
B. hyohysenteriae



Demonstration of
***B. hyodysenteriae* by FISH**

Intestinal diseases and differential diagnostic problems





PCV2 enteritis

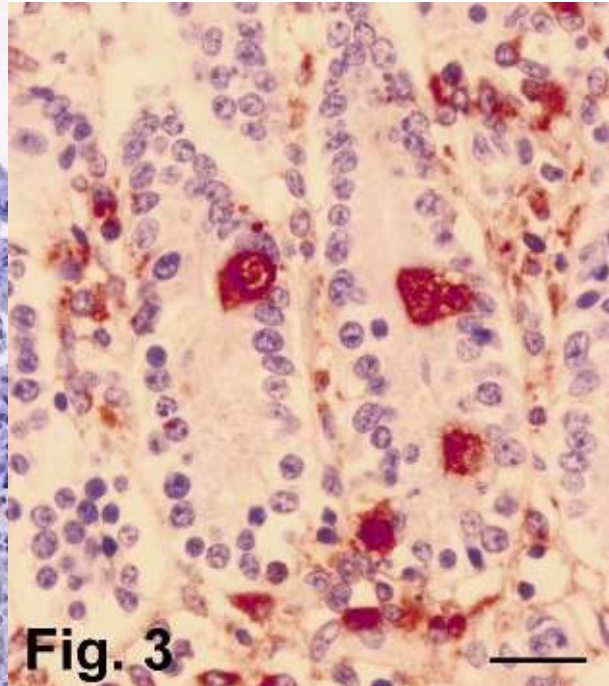
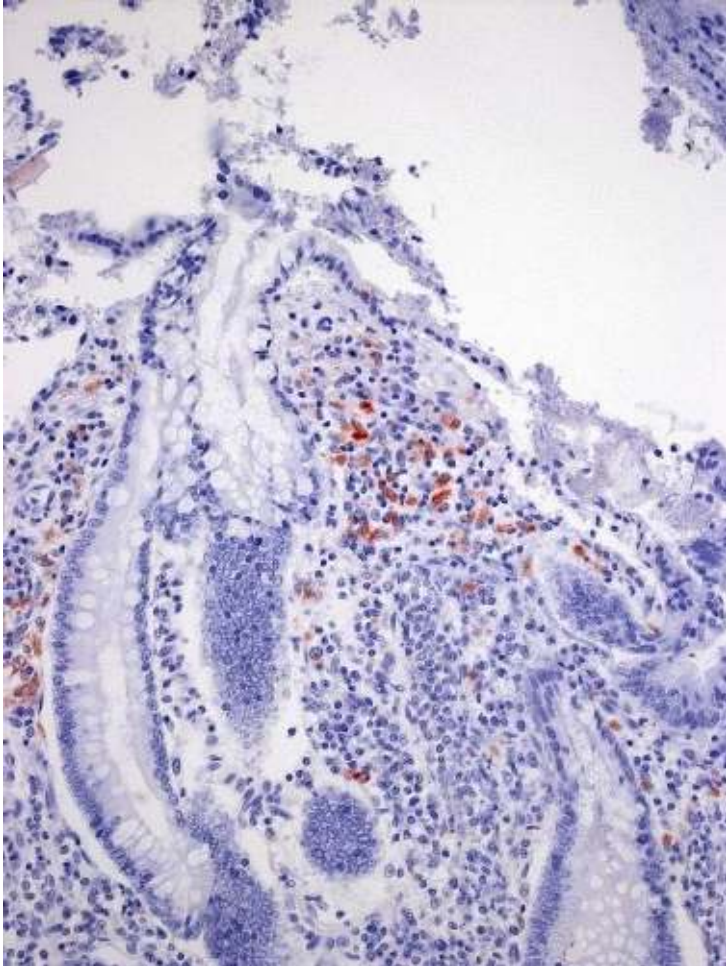
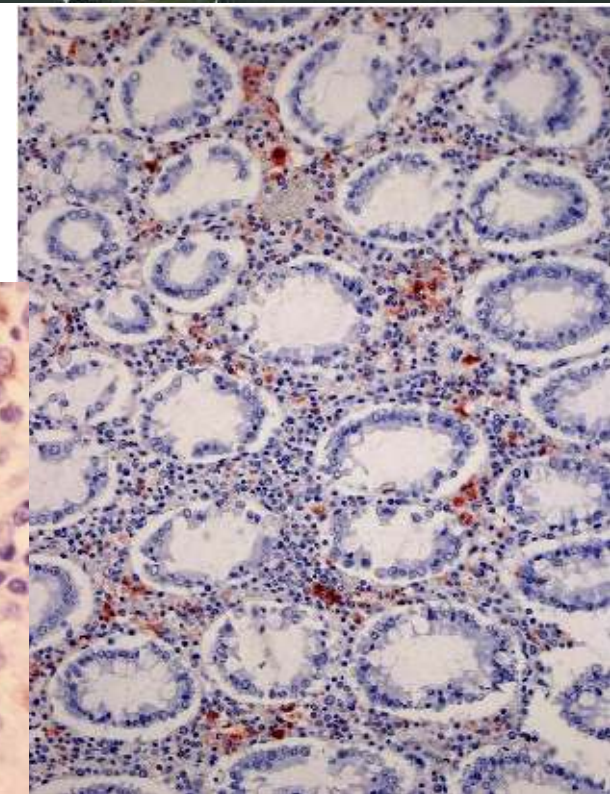


Fig. 3



Demonstration of PCV2 by IHC

Detection of large intestinal pathogens in pigs by in situ methods

Intestines from 140 pigs submitted for routine laboratory examination with suspicion of spirochaete associated diarrhoea/colitis. FISH and IHC.

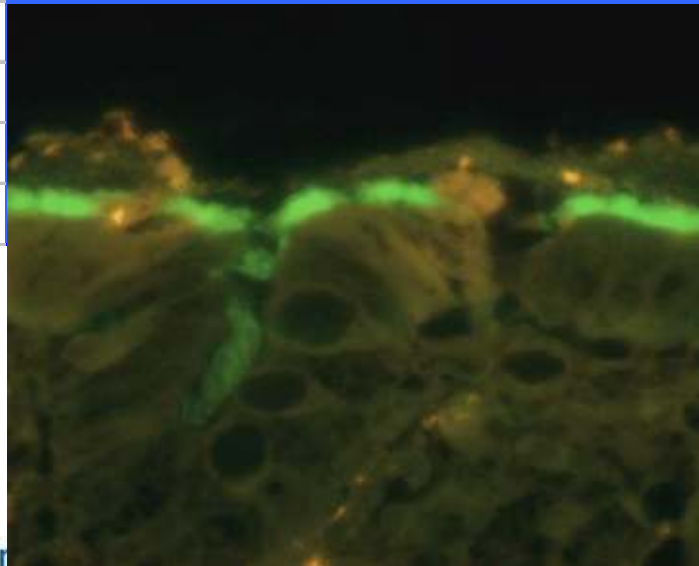
Demonstration of mono- and polyinfections in pigs

	<i>L.intra</i>	<i>Bhy+Bpil</i>	<i>B.hyo</i>	<i>B.pilo</i>	<i>B.inter</i>	B. inn	B. mur	PCV2
<i>L.intra</i>	28	2	7	1	1	2	3	5
<i>B.hyo</i>			22	4		2		
<i>B.pilo</i>				3		1		2
<i>B.inter</i>					2	1		
B. inn						3		2
B. mur							3	
PCV2								14

In situ detection of large intestinal pathogens in pigs

Demonstration of mono- and polyinfections in pigs

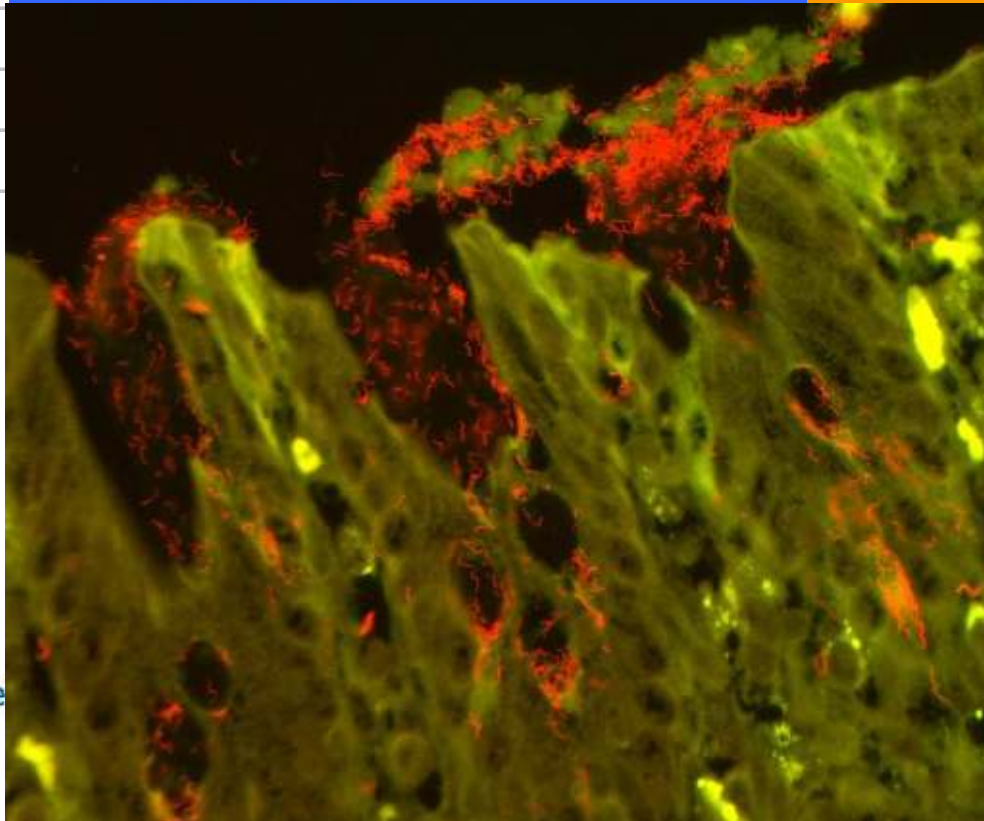
	<i>L.intra</i>	<i>Bhy+Bpil</i>	<i>B.hyo</i>	<i>B.pilo</i>	<i>B.inter</i>	<i>B. inn</i>	<i>B. mur</i>	PCV2
<i>L.intra</i>	28	2	7	1	1	2	3	5
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<i>B.pilo</i>				3		1		2
<i>B.inter</i>								
<i>B. inn</i>								
<i>B. mur</i>								
PCV2								



In situ detection of large intestinal pathogens in pigs

Demonstration of mono- and polyinfections in pigs

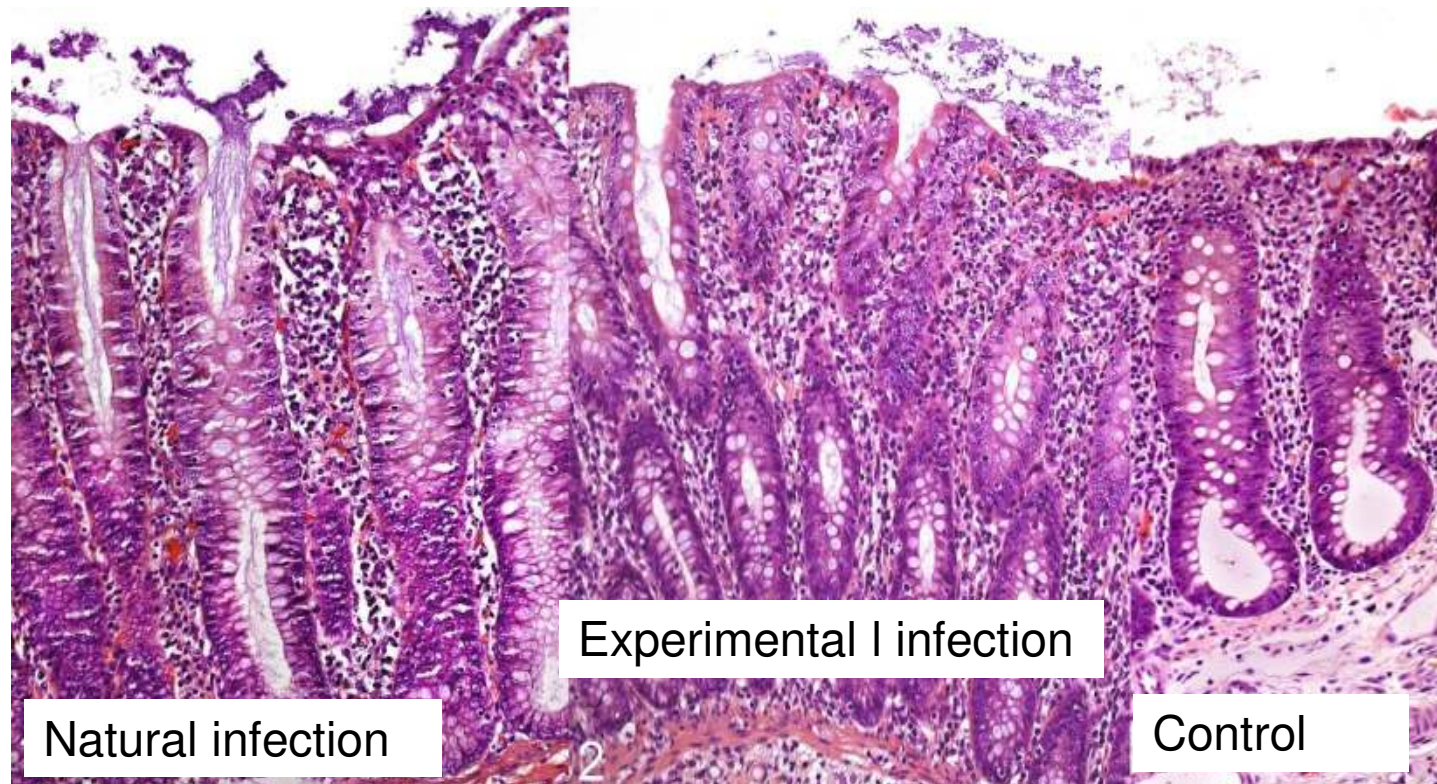
	<i>L.intra</i>	<i>Bhy+Bpil</i>	<i>B.hyo</i>	<i>B.pilo</i>	<i>B.inter</i>	B. inn	B. mur	PCV2
<i>L.intra</i>	28	2	7	1	1	2	3	5
<i>B.hyo</i>			22	4		2		
<i>B.pilo</i>			3		1		2	
<i>B.inter</i>					2	1		
B. inn					3		2	
B. mur							3	
PCV2								



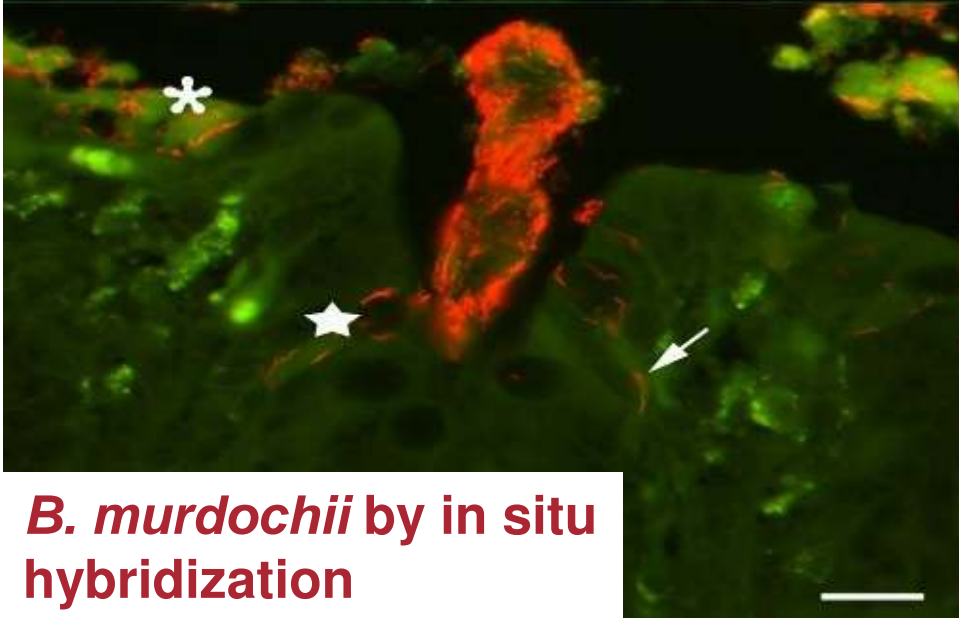
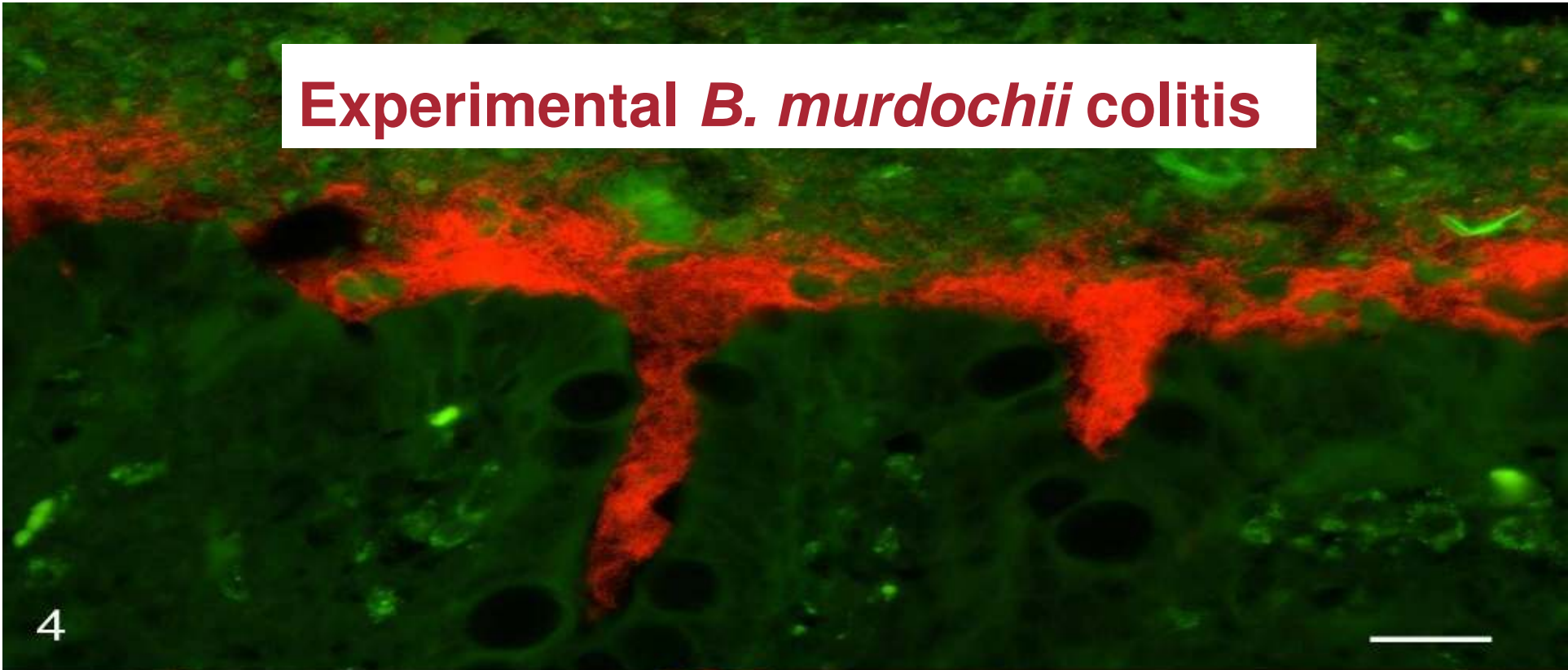
Experimental *B. murdochii* colitis

Two out of 8 pigs showed catarrhal colitis 3 weeks after challenge with *B. murdochii*.

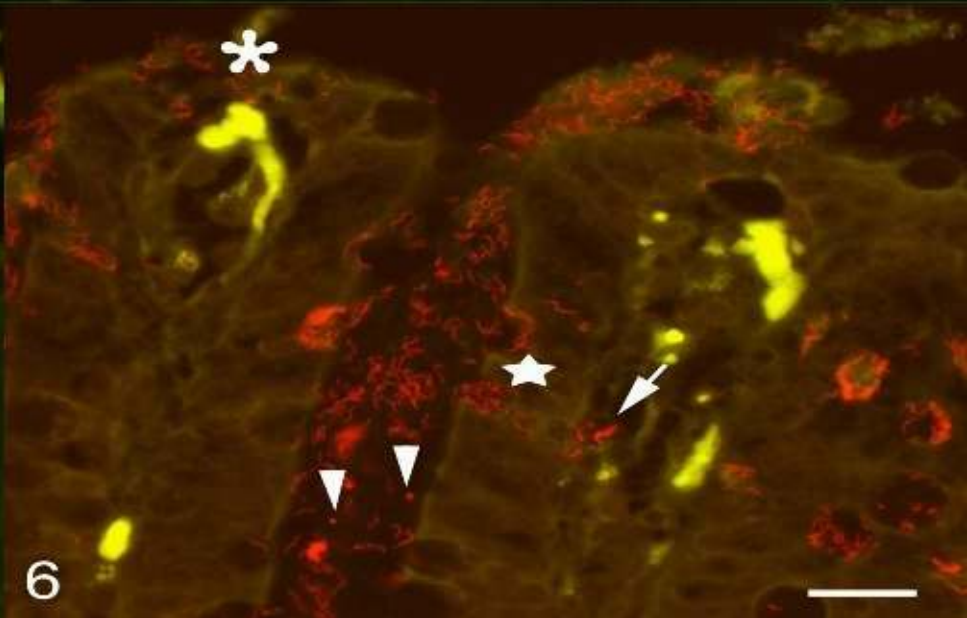
Diarrhoea was not observed.



Experimental *B. murdochii* colitis



B. murdochii by in situ hybridization



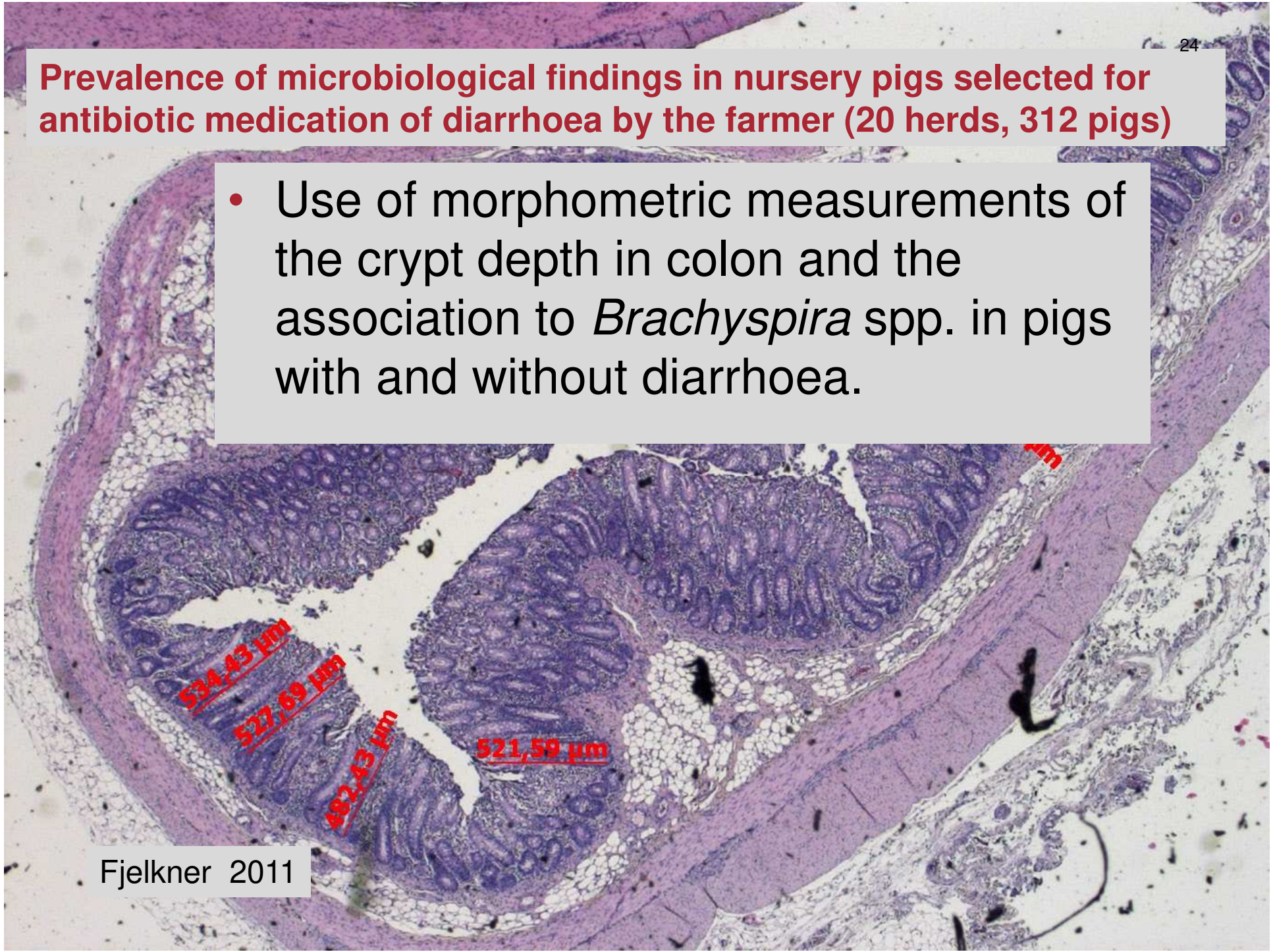
Prevalence of microbiological findings in nursery pigs selected for antibiotic medication of diarrhoea by the farmer (20 herds, 312 pigs)

Pig prevalences (%)

• <i>L. intracellularis</i>	19
• PCV2	4
• <i>Brachyspira spp</i>	40
<i>B. pilosicoli</i>	7
<i>B. intermedia</i>	10
<i>B. innocens</i>	16
<i>B. murdochii</i>	21
• Hemolytic <i>E. coli</i>	15

Prevalence of microbiological findings in nursery pigs selected for antibiotic medication of diarrhoea by the farmer (20 herds, 312 pigs)

- Use of morphometric measurements of the crypt depth in colon and the association to *Brachyspira* spp. in pigs with and without diarrhoea.



Prevalence of microbiological findings in nursery pigs selected for antibiotic medication of diarrhoea by the farmer (20 herds, 312 pigs)

- An increased thickness of the colonic mucosa was associated with demonstration of *Brachyspira* spp. (*B. pilosicoli*, *B. intermedia*, *B. innocens* and/or *B. murdochii*) ($p=0.04$).
- The mean thickness of the colonic mucosa in pigs positive for *Brachyspira* spp. was 529.5 μm compared to 501.4 μm in *Brachyspira* spp. negative pigs.
- Increased thickness of the colonic mucosa was associated with small pig size ($p = 0.05$) and increasing age ($p= 0.03$).

Colon morphometry in healthy and diseased pigs

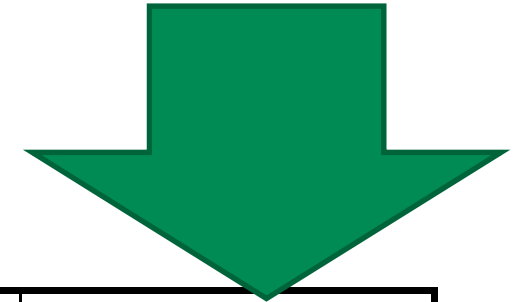
- Svinedysenteri: 800-1200 μm
- Brachyspira positive svin 530 μm
- Brachyspira negative svin 501 μm
- Colon fra raske svin: 378 μm
kontroldyr fra podningsforsøg
- Lindholmgrise 250 μm

482,43

521,59 μm

Porcine necrotizing enterocolitis

Gross lesions and detection of the agents: *L. intracellularis* and PCV2
in 64 pigs with enteritis



Law(n=28)	Law+PCV2 n=6)	PCV2 (n=23)	None (n=7)
Uncomplicated PE (15) Necrotizing enterocolitis (10) PHE (3)	Uncomplicated PE (3) Necrotizing enterocolitis (3)	Necrotizing enterocolitis (12) Other lesions (11)	Necrotizing enterocolitis (6) Other lesions (1)

Porcine necrotizing enterocolitis

- Necropsy
Severe, acute necrotizing enterocolitis was found in both animals while all other organs appeared normal.

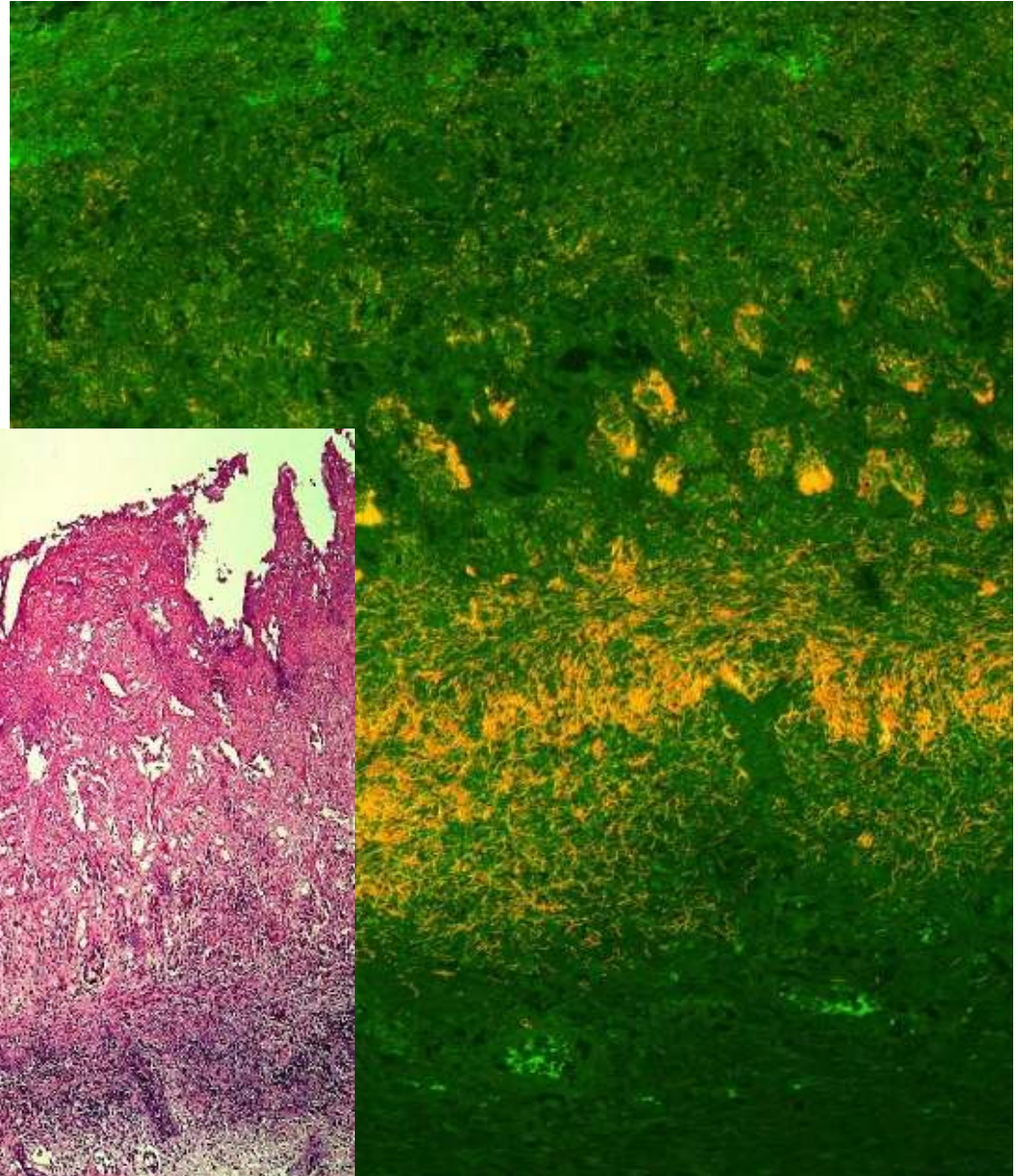


Porcine *Fusobacterium necrophorum* necrotizing enterocolitis

29

Histopathology:

Acute to subacute
necrotizing enterocolitis.
High number of slender rods
observed.



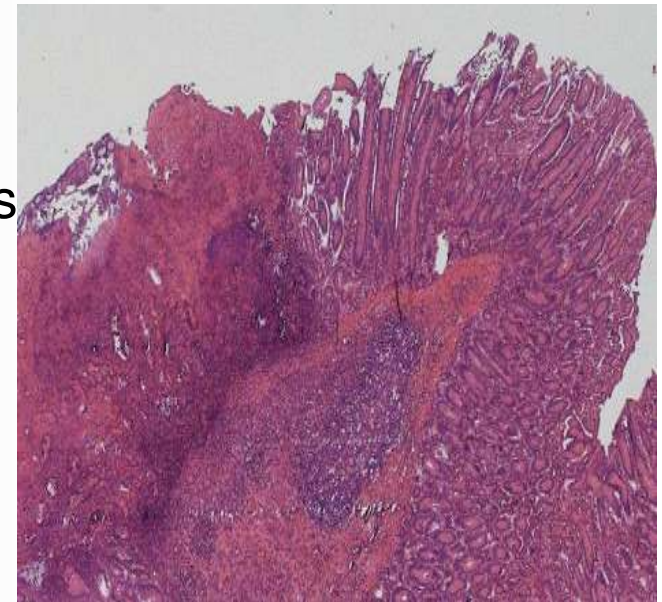
Porcine *Fusobacterium necrophorum* necrotizing enterocolitis

30

Retrospective FISH study on 84 intestines from pigs with suspicion of *L. intracellularis* infection (33+/51-)

Necrotic mucosa associated with *F. necrophorum* colonization detected in 28 pigs (33%)

- *L. intracellularis* +: 18 cases (55%)
- *L. intracellularis* -: 10 cases (20%)
 - Including 4 severely affected cases



Knowing how *F. necrophorum* looks like: Nine out nine positive! Age: 6 days to 6 weeks

Intestinal diseases in pigs

Conclusion

- The results suggest that *F. necrophorum* in some cases may act as a primary intestinal pathogen.
- Large scale studies to be initiated to investigate the importance of *F. necrophorum* as an intestinal pathogen.
- *B. intermedia*, *B. murdochii* and *B. innocens* should be regarded as low pathogenic for pigs.

Intestinal diseases in pigs

Conclusions

- The diagnostic importance, however, of culturing *B. intermedia*, *B. murdochii* and *B. innocens* from feces only is uncertain, as the method is not quantitative!
- Diagnostic pathology is difficult – not all cases are straight forward!
- Application of different in situ detection methods is a requirement for research as well as diagnostic laboratories.



Diarrhoea in fattening pigs

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Thank you for your attention